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Atty's Docket No.
2212.135/00

EXPRESS MAIL CERTIFICATION

"Express" Mail label number: EL435572490 US

(A) Date of Deposit: November 12, 1999

I hereby certify that this transmittal letter and the papers and fees identified in this transmittal letter as being transmitted herewith are being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 C.F.R. 1.10 on the date indicated at (A) above and are addressed to the Assistant Commissioner of Patents, Washington, D.C. 20231

Name of Person mailing the above: Kathleen D. Monical

Signature of Person mailing the above item

TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (D0/EO/US)

International Application No	: PCT/NL98/00259
International Filing Date	: 11 May 1998 (11.05.98)
Priority Date Claimed	: 12 May 1997 (12.05.97)
Title of Invention	: Method and Construct for Inhibition of Cell Migration
Applicant(s) for DO/EO/US	: Paulus Hubertus Quax and Johan Hendrijus Verheijen

Applicant herewith submits to the United States Designed/Elected Office (DO/EO/US) the following items under 35 U.S.C. 371:

1. ✓ This express request to immediately begin national examination procedures (35 U.S.C. 371(f).

2. ✓ The U.S. National Fee (35 U.S.C. 371(c)(1) and other fees as follows:

TOTAL CLAIMS 25 - 20 =	CLAIMS OVER 20 5	RATE X $18 =	TOTAL FEES FOR CLAIMS OVER 20 90.00
NUMBER OF INDEPENDENT CLAIMS 3 - 3 =	CLAIMS OVER 3 --	RATE X $78 =	TOTAL FEES FOR INDEPENDENT CLAIMS OVER 3 --
MULTIPLE DEPENDENT CLAIM(S) PRESENT No		RATE $260 per APPLN.	FEE MULTIPLE DEPENDENT CLAIM(S) --
BASIC NATIONAL FEE (37CFR 1.492(a)(1)-(4)): __ International preliminary Examination fee paid to USPTO (37 CFR 1.482) = $670.00 __ No International preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2) = $760.00 __ Neither international preliminary examination fee (37 CFR 1.482) nor International Search fee (37 CFR 1.445(a)(2) paid to USPTO = $970.00 __ International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)(2) to (4) = $96.00 ✓ Filing with an EPO or JPO search report = $840.00			840.00
Surcharge of $130 for furnishing the national fee or oath or declaration 20 mos. from the earliest claimed priority date (37 CFR 1.482(e)).			130.00
		TOTAL OF ABOVE CALCULATIONS	1,060.00
Reduction by ½ for filing by small entity			530.00
		SUBTOTAL	530.00
Process fee of $130 for furnishing the English translation later than 20 mos from the earliest claimed priority date (37 CFR 1.482(f))			
		TOTAL NATIONAL FEE	530.00
Fee for recording the enclosed assignment			
		TOTAL FEES ENCLOSED	

a. __ A check in the amount of $ to cover the above fees is enclosed.
b. __ Please charge my Deposit Account No. in the amount of $ to cover the above fees. A duplicate copy of this sheet is enclosed.
c. __ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. . A duplicate copy of this sheet is enclosed.

3. A copy of the International Application as filed (35 U.S.C. 371(c)(2)
   a. _ is transmitted herewith (required only if not transmitted by the International Bureau).
   b. _ is not required, as the application was filed in the United States Receiving Office.
   c. ✓ has been transmitted by the International Bureau.

4. _ A translation of the International Application into English.

5. Amendments to the claims of the International Application under PCT Article 19
   a. _ are transmitted herewith (required only if not transmitted by the International Bureau).
   b. _ have been transmitted by the International Bureau.

6. _ A translation of the amendments to the claims under PCT Article 19

7. _ An oath or declaration of the inventor [35 U.S.C. 371(c)(4)]

8. _ A translation of the Annexes to the International Preliminary Examination Report under PCT Article 36 (35. U.S.C. 371(c)(5)).

Other document(s) or information included:
9. ✓ Preliminary Amendment
10. _ An assignment document for recording. Please mail the recorded assignment document to the undersigned.
11. ✓ The above checked items are being transmitted
   a. _ before the 18th month publication.
   b. _ after publication and the Article 20 communication but before 20 months from the priority date.
   c. _ after 20 months (surcharge and/or processing fee included).
      Note: Petition to revive (37 CFR 1.137(a) or (b)) is necessary if 35 U.S.C. 371 requirements submitted after 20 months and no proper demand for International Preliminary Examination was made by 19 months from the earliest claimed priority date.
   e. ✓ by 30 months and a proper demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
   f. _ after 30 months (surcharge and/or processing fee included).
      Note: Petition to revive (37 CFR 1.137(a) or (b)) is necessary if 35 U.S.C. 371 requirements submitted after 32 months and a proper demand for International Preliminary Examination was made by 19 months from the earliest claimed priority date.

12. At the time of transmittal, the time limit for amending claims under Article 19
   a. ✓ has expired and no amendments were made.
   b. _ has not yet expired.

13. _ Certain requirements under 35 U.S.C. 371 were previously submitted by the applicant on _____ namely:

Please direct all communications in connection with this application to the undersigned at

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New York, N.Y. 10005

Bruce S. Londa (33,331)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Atty's Docket No: 2212.135/00

Applicant(s) : Paulus Hubertus Quax et al.

Filed : Concurrently herewith

For : Method and Construct for Inhibition of Cell Migration

PRELIMINARY AMENDMENT

Hon. Assistant Commissioner of Patents
Washington, D.C. 20231

Dear Sir:

Prior to examination, please amend the application as follows:

IN THE CLAIMS

Please amend the following claims:

20. (amended) A process for preventing local proteolytic activity, extracellular matrix degradation, cell migration, cell invasion, or tissue remodeling, comprising transfecting or transducing the cells involved or cells in their environment with a recombinant nucleic acid molecule as claimed in [any one of the preceding Claims] Claim 1 to obtain local expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule.

21. (amended) A process for producing a hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function, comprising transfecting or

transducing mammalian cells with a recombinant nucleic acid molecule as claimed in [any one of Claims 1 to 19] Claim 1 to obtain expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule, and optionally recovering the hybrid polypeptide or protein produced.

Please add the following new claims:

22. A process for preventing local proteolytic activity, extracellular matrix degradation, cell migration, cell invasion, or tissue remodeling, comprising transfecting or transducing the cells involved or cells in their environment with a recombinant nucleic acid molecule as claimed in Claim 18 to obtain local expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule.

23. A process for preventing local proteolytic activity, extracellular matrix degradation, cell migration, cell invasion, or tissue remodeling, comprising transfecting or transducing the cells involved or cells in their environment with a recombinant nucleic acid molecule as claimed in Claim 19 to obtain local expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule.

24. A process for producing a hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function, comprising transfecting or transducing

mammalian cells with a recombinant nucleic acid molecule as claimed in Claim 18 to obtain expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule, and optionally recovering the hybrid polypeptide or protein produced.

25. A process for producing a hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function, comprising transfecting or transducing mammalian cells with a recombinant nucleic acid molecule as claimed in Claim 19 to obtain expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule, and optionally recovering the hybrid polypeptide or protein produced.

IN THE ABSTRACT

Please add the following abstract:

--Abstract of the Disclosure

A recombinant nucleic acid molecule comprising a vector useful for transfection or transduction of mammalian cells, wherein said vector contains a nucleic acid insertion encoding an expressible hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function. The domain with a binding function may comprise a receptor binding domain, and the domain with an effector function may have enzymatic activity, in particular protease inhibitor activity. The vector may be a viral (e.g. adenovirus or retrovirus) or non-viral vector useful for transfection or

transduction of mammalian cells. The nucleic acid insertion encoding an expressible hybrid polypeptide or protein may be under the control of a cell- or tissue-specific promoter. A process for preventing local proteolytic activity, extracellular matrix degradation, cell migration, cell invasion, or tissue remodeling, comprising transfecting or transducing the cells involved or cells in their environment with the recombinant nucleic acid molecule to obtain local expression of the hybrid polypeptide or protein encoded thereby. A process for producing the hybrid polypeptide or protein by transfecting or transducing mammalian cells with the recombinant nucleic acid molecule to obtain expression and optionally recovering the hybrid polypeptide or protein produced.--

REMARKS

The above amendments were made to remove multiple dependent claims. Early and favorable consideration is earnestly solicited.

Respectfully Submitted,

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Title: Method and Construct for inhibition of cell migration

FIELD OF THE INVENTION

The invention is in the field of therapeutic means and therapeutic methods for treatment of diseases in which cell migration and/or tissue remodeling occurs. Furthermore, the invention is in the field of biotechnology, in particular recombinant DNA technology and gene therapy.

BACKGROUND OF THE INVENTION

Migration of cells is an essential step in many physiological and pathological processes in which tissue remodeling occurs, such as tumor metastasis, wound healing, restenosis, angiogenesis or rheumatic arthritis. Migrating cells have to pass through the surrounding extracellular matrix. Limited proteolytic degradation of the components of the extracellular matrix is often seen during cell migration. To mediate this cell migration migrating cells produce, or recruit from their direct environment, proteolytic enzymes, such as plasminogen activators, metalloproteinases or elastases. Induction of cell migration e.g. during tumor metastasis or wound healing often correlates with the induction of the production of these enzymes.

Although the involvement of proteolytic enzymes in cell migration under pathophysiological conditions is well accepted, little attempts have been made to inhibit cell migration by inhibiting these proteolytic enzymes. A possible explanation for the limited use of protease inhibitors is the fact that these proteolytic enzymes are involved in many processes both pathological and physiological (including fibrinolysis, wound healing, growth factor activation etc.) and that inhibition of these protease systems by systemically applied protease inhibitors might have either strong side effects or may lead to a diffusion or clearance of the inhibitory compounds without having a strong effect on the local cell migration processes.

Another problem in the use of protease inhibitors to interfere in cell migration and tissue remodeling is that proteases mediating these processes can bind to receptors at the cell surface. In this way the proteolytic enzymes might be active locally in a pericellular microenvironment where they are protected against the action of the present inhibitors.

It has been disclosed that conjugates between the receptor binding part of u-PA (the aminoterminal fragment or ATF) and urinary trypsin inhibitor produced in vitro, inhibit migration of tumor cells in vitro (Kobayashi, Gotoh, Hirashima, Fujie, Sugino and Terao, Inhibitory effect of a conjugate between human urokinase and urinary trypsin inhibitor on tumor cell invasion in vitro. J. Biol. Chem. (1995) 270, 8361-8366). The conjugate these authors have used is made synthetically by mixing the isolated ATF fragments with the trypsin inhibitor.

Recently it has been disclosed that these conjugates also can be produced recombinantly (WO 97/25422).

A comparable construct consisting of a receptor binding u-PA fragment and its inhibitor PAI-2, to be produced recombinantly in yeast, has been described to inhibit tumor cell migration in WO 92/02553 (PCT/GB91/01322). In this way they have made a protease inhibitor that can bind to a specific receptor at the cell surface, the urokinase receptor, and this inhibitor can inhibit cell migration (in vitro). As to the use of these constructs in vivo, a problem is the application to and the prolonged presence at the site of desired action in vivo.

SUMMARY OF THE INVENTION

This invention provides a recombinant nucleic acid molecule comprising a vector useful for transfection or transduction of mammalian, e.g. human, cells, wherein said vector contains a nucleic acid insertion encoding an expressible hybrid polypeptide or protein which comprises a

domain with a binding function and a domain with an effector function. Herein, the domain with a binding function preferably comprises a receptor binding domain, and the domain with an effector function preferably has enzymatic activity, most preferably protease inhibitor activity.

Preferably, the receptor binding domain is selected from the group consisting of urokinase receptor binding domain of urokinase, receptor binding domain of epidermal growth factor, receptor associated protein that binds to LDL Receptor related protein (α_2-macroglobulin receptor) and VLDL Receptor.

Preferably, the domain with an effector function has protease inhibitor activity and comprises a protease inhibitor or active part thereof, said protease inhibitor being selected from the group consisting of (bovine) pancreatic trypsin inhibitor, (bovine) splenic trypsin inhibitor, urinary trypsin inhibitor, tissue inhibitor of matrix metalloproteinase 1, tissue inhibitor of matrix metalloproteinase 2, tissue inhibitor of matrix metalloproteinase 3, and elastase inhibitor. The domain with an effector function may comprise (an active part of) two or more different protease inhibitors, or two or more copies of (an active part of) a protease inhibitor, or both.

Preferably, the vector is selected from the group consisting of viral and non-viral vectors useful for transfection or transduction of mammalian cells. The vector may be an adenovirus vector or a retrovirus vector useful for transfection or transduction of human cells.

The nucleic acid insertion encoding an expressible hybrid polypeptide or protein may be under the control of a cell- or tissue-specific promoter, such as an endothelial cell-specific promoter, or a vascular smooth muscle cell-specific promoter, or a liver-specific promoter.

This invention furthermore provides a process for preventing local proteolytic activity, extracellular matrix degradation, cell migration, cell invasion, or tissue

remodeling, comprising transfecting or transducing the cells involved or cells in their environment with a recombinant nucleic acid molecule as defined herein to obtain local expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule.

Also, this invention provides a process for producing a hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function, comprising transfecting or transducing mammalian cells with a recombinant nucleic acid molecule as defined herein to obtain expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule, and optionally recovering the hybrid polypeptide or protein produced.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 schematically depicts the plasmids pCRII-uPA (left) and pCRII-ATF (right).

Figure 2 schematically depicts the plasmid pCRII-ATF-BPTI.

Figure 3 schematically depicts the plasmid pMAD5-ATF-BPTI.

Figure 4 shows the results of proteolytic matrix degradation experiments.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to the use of hybrid proteins in which a receptor binding domain is linked to a functional protein in order to induce a local action of this protein and to prevent systemic effects and/or diffusion. In particular this invention relates to such hybrid proteins that might be produced by a subset of cells as target cells after transfection or transduction with expression vectors. More specifically the invention relates to the use of such expression vectors, coding for hybrid proteins consisting of a receptor binding domain and a protease inhibitor domain, for the prevention of cell migration and tissue remodeling by

inhibition of proteases at the surface of migrating or invading cells.

The method and construct described in the present invention can be applied as therapy in diseases in which cell migration and/or tissue remodeling occurs.

The present invention addresses the solution of several negative aspects involved in the above described use of inhibitors according to the prior art:

- High local concentrations of hybrid proteins in the direct environment of the target cells can be obtained by production of the protein by the migrating cells themselves or cells in their immediate environment. This production can be induced by transfection or transduction of a certain subset of the cell population with a suitable vector encoding the hybrid protein. For this purpose, one may use recombinant adenoviral vectors, retroviral vectors, plasmid vectors, etc.
- Diffusion of the inhibitor and systemic side effects are prevented by binding the hybrid protein (by its receptor binding domain) to the cell surface of the target cell. Local expression of this hybrid protein also contributes to the reduction of systemic side effects, while the negative effect of diffusion of the protein is reduced by the production at the site where action is required. The local expression of the hybrid protein in specific subpopulations of cells, e.g. endothelial cells prone to migrate during angiogenesis, can be enhanced using cell type-specific or tissue-specific expression vectors, in which the expression of the protein is under control of a promoter with cell type-specific or tissue-specific regulatory elements.
- Binding of a protease inhibitor to a cell surface receptor can locate the inhibitor close to its molecular target, the cell surface bound proteolytic enzyme. Local inhibition of the proteolytic activity in the pericellular microenvironment may be achieved in this way.
- Binding of a protease inhibitor to a cell surface receptor for a proteolytic enzyme, such as the urokinase

receptor, may have an additional inhibitory effect. It prevents the binding of the proteolytic enzyme to its receptor, and thus strongly reduces the action of this enzyme as has been shown for blocking the binding of u-PA to its receptor which can strongly inhibit cell migration.

Hybrid proteins, for which the expression vectors (e.g. adenoviral or retroviral expression vectors) contain the encoding DNA sequences, might contain a region that binds to a cell surface receptor and that is not subsequently internalized. Receptor binding domains that can be used for this purpose are e.g. the u-PAR binding domain of urokinase plasminogen activator, the receptor binding domain of epidermal growth factor, the receptor associated protein (RAP) that binds to the LDL-R related protein (LRP), also called α_2-macroglobulin receptor, and the VLDL-receptor.

The inhibitor part of the encoded hybrid protein might consist of various protease inhibitors such as bovine pancreatic trypsin inhibitor, also called aprotinin or Trasylol®, other trypsin inhibitors such as urinary trypsin inhibitor, inhibitors for matrix-degrading metalloproteinases such the tissue inhibitors of metalloproteinases TIMP-1, TIMP-2 and TIMP-3, or variants thereof. Also inhibitors for other proteases like elastase are very suitable for being incorporated into the expression vector containing the DNA sequences encoding the hybrid proteins. Multiple copies of the DNA sequences encoding the functional protein part of the hybrid protein e.g. the inhibitor part, or combinations of different inhibitors or derivatives thereof might be incorporated into the DNA construct in the expression vector.

Another very attractive possibility would be to use such an expression vector encoding hybrid receptor binding protein to apply any functional protein that should exert its action in the local environment of the target cell, e.g. a protease involved in the activation of a growth factor or an other e.g. vasoregulatory component.

The action of the functional protein or protein domains of the hybrid protein is localized to the direct microenvironment of the target cells by binding of the receptor binding domain to a receptor at the surface of the target cells. Production of the hybrid protein in the direct environment of the target cells or even by the target cells themselves can be obtained by transfection or transduction of these cells by the use of expression vectors that might be based on a non-viral or an adeno- or retroviral vector system. Expression in specific cell or tissue types might be achieved by the use of specific promoter elements in the expression vectors. For example, for endothelial cell-specific expression (elements of) the promoter region of the human or murine pre-pro-endothelin gene (HUMEDN1B and MMU07982, respectively, GENBANK) can be used, for vascular smooth muscle cell-specific expression (elements of) the promoter region of the human vascular smooth muscle α-actin gene (HUMACTSA, GENBANK) can be used, and for liver-specific expression the promoter of the human albumin gene (HUMALBGC, GENBANK) can be used.

Local delivery of these vectors might be obtained using various commonly used methods, including catheters, topically applied gels containing the vectors or targeted delivery systems. For site-specific delivery to the vessel wall, e.g. to prevent restenosis and vessel wall remodeling after angioplasty, special catheters can be used. At the moment double balloon catheters, channeled balloon catheters, multiple needle catheters and balloon catheters coated with a vector containing a hydrogel are being used for vessel wall-specific delivery. Other ways to deliver the vectors directly into the vessel wall are the use of stents coated with vector containing coatings, topical application of vector containing hydrogels to the outside of the blood vessel or ex vivo delivery directly into the blood vessel during trans-plantation surgery. Ex vivo transduction of proliferating cells using retroviral vectors followed by a reinjection may

also be used to deliver the vector constructs at the site where their action is required.

The present application will be described hereinafter in further detail, while referring to the following examples. It is to be noted that these examples merely serve to illustrate the invention, not to restrict it.

EXAMPLE 1

An expression plasmid encoding the aminoterminal fragment of urokinase plasminogen activator (u-PA), amino acids 1-138, hereafter referred to as ATF, can be constructed by deleting the DNA sequences encoding amino acids 139 till 401 in an expression plasmid for the full length u-PA using a polymerase chain reaction (PCR) with the following oligonucleotides: 5'-cccgggctttttccatctgcgcagtc-3' and 5'-agggtcaccaaggaagagaatggc-3'. After amplification by PCR the newly formed DNA fragment can be circularized by ligation to restore the circular character of the expression plasmid. In this way an expression plasmid encoding the ATF and the C terminal last 11 amino acid residues including the stop codon can be constructed.

The sequence of the thus formed DNA construct encoding the u-PA ATF fragment then is determined and compared with the predicted sequence as a control for possible mutations introduced during the construction procedure.

The construction pCRII-ATF from pCRII-uPA using PCR is shown in Figure 1. In figure 1, the area indicated between the lines was removed during the PCR amplification, resulting in the ATF plasmid. The plasmid pCRII-uPA is shown to the left, plasmid pCRII-ATF to the right.

EXAMPLE 2

DNA fragments encoding amino acid residues 36-93 of bovine pancreatic trypsin inhibitor (BPTI) and the homologous amino acid residues of bovine spleen trypsin inhibitor (BSTI)

can be isolated by performing a PCR reaction on genomic DNA isolated for bovine aortic endothelial cells using the following oligonucleotides: 5'-*tcgcga*cctgacttctgcctagagc-3' covering nucleotides 2509 to 2533 (with modifications, indicated in *italics*, in the 5' region of the oligonucleotide to introduce a NruI site (underlined) for cloning purposes) of the BPTI gene according to the published sequence (GENBANK, BTBPTIG), and nucleotide 2442 to 2462 of the BSTI gene according to the published sequence (GENBANK, BTBSTIG) and 5'-*ggtcacc*cagggcccaatattaccacc-3' covering nucleotides 2677 to 2704 of the BPTI gene and 2610 to 2636 of the BSTI gene (modified in the indicated nucleotides (*italics*) to introduce a BstEII and a SspI site respectively (underlined)). The amplified DNA fragments then were cloned into an appropriate plasmid vector, pCRII or pUC13, and then the exact sequence of the amplified DNA fragments in the isolated clones was analyzed to differentiate between BPTI and BSTI which have a very high degree of homology.

EXAMPLE 3

The DNA fragment encoding amino acids 1 to 207 of the human tissue inhibitor of metalloproteinase type 1 is isolated by performing a reverse transcriptase polymerase chain reaction on total RNA isolated from human foreskin fibroblasts by using the following oligonucleotides 5'-agagagacaccagagaacccaccat-3' covering nucleotides 41 to 65 of the human TIMP-1 cDNA (according to the sequence in GENBANK HSTIMPR) and 5'-tcattgtccggaagaaagatgggag-3' covering nucleotides 740 till 755. The amplified DNA fragment was cloned into an appropriate host vector, pUC13, and then the exact sequence of the amplified DNA fragment in the isolated clones was analyzed.

EXAMPLE 4

For construction of a recombinant adenovirus containing sequences encoding the ATF.BPTI hybrid protein,

this sequence is cloned in the adenoviral vector construction adapter and expression plasmid pMAD5. This plasmid contains part of the wildtype adenovirus type 5 DNA sequences, a Major Late Promoter (MLP), and a poly-adenylation (polyA) signal and can be used as either an expression vector or a shuttle vector to construct a recombinant adenovirus. This plasmid was derived from plasmid pMLP10 as follows. First pMLP10-lin was constructed by insertion of a synthetic DNA fragment with unique sites for the restriction endonucleases MluI, SplI, SnaB1, SpeI, AsuII and MunI into the HindIII site of pMLP10. Subsequently, the adenovirus BglII fragment spanning nt 3328 to 8914 of the Ad5 genome was inserted into the MunI site of pMLP10-lin. Finally, the SalI-BamHI fragment was deleted to inactivate the tetracycline resistance gene, resulting in plasmid pMAD5. To clone the ATF.BPTI sequence into the pMAD5 plasmid between the MLP promoter and the polyA signal the following strategy has been followed.

Starting from a pCRII plasmid in which a 1373 base pair fragment of the uPA cDNA was cloned, a PCR reaction with the oligonucleotides 5'-*cccggg*cttttttccatctgcgcagtc-3' (SmaI site underlined and nucleotides changed in *italics*) and 5'-ag*ggtcacc*aaggaagagaatggc-3' (BstEII site underlined and nucleotides changed in *italics*) was performed as described in example 1 to make a pCRII-ATF plasmid (see figure 1). Subsequently this pCRII-ATF plasmid was digested with the restriction enzymes SmaI and BsteII. In parallel the pCRII-BPTI plasmid was digested with the restriction enzymes NruI and BsteII and the BPTI containing fragment was cloned into the pCRII-ATF plasmid (see figure 2). The construction pCRII-ATF-BPTI is shown in Fig. 2.

In a next step the ATF-BPTI sequence was cloned into pMAD5. This was done by digestion of the pCRII-ATF-BPTI plasmid with the restriction enzymes EcoRV and SpeI, isolation of the ATF-BPTI encoding DNA fragment and cloning of this fragment into the SnaBI and SpeI digested pMAD5

plasmid. The cloning was tested by restriction analysis and sequence analysis.

The pMAD5-ATF-BPTI shuttle vector for the construction of ATF-BPTI adenoviral vector is shown in Figure 3.

EXAMPLE 5

In a similar way as described in example 4 for pMAD5-ATF-BPTI a plasmid containing the BSTI-gene (pMAD5-ATF-BSTI) was constructed using the pCRII-BSTI plasmid instead of the pCRII-BPTI plasmid.

EXAMPLE 6

For construction of a recombinant adenovirus containing sequences encoding the ATF-TIMP1 hybrid protein, this sequence is cloned in the pMAD5 expression plasmid. This plasmid contains part of the wildtype adenovirus type 5 DNA sequences, a Major Late Promoter (MLP), and a polyadenylation (polyA) signal and can be used as either an expression vector or a shuttle vector to construct a recombinant adenovirus. To clone the ATF-TIMP1 sequence into the pMAD5 plasmid between the MLP promoter and the polyA signal, the following strategy has been followed.

Starting from a pCRII plasmid in which a 1373 base pair fragment of the uPA cDNA was cloned, a PCR reaction with the oligonucleotides 5'-cccgggctttttttccatctgcgcagtc-3' and 5'-agggtcaccaaggaagagaatggc-3' was performed as described in example 1 to make a pCRII-ATF plasmid (see figure 1). Subsequently this pCRII-ATF plasmid was digested with the restriction enzymes SmaI and BsteII.

In parallel a fragment of the cDNA of TIMP1 in pUC13-TIMP1 encoding amino acid residues 1 to 184 of the mature protein, but lacking the signal peptide and the stop codon, was amplified using the following oligonucleotides 5'-tcgcgatgcacctgtgtcccacc-3' and 5'-ggtcacccaaatattggctatgtgggaccgcaggg-3'. These oligonucleotides contain recognition sites for the restriction

enzymes NruI (first oligonucleotide, underlined) and BstEII and Sspl respectively (second oligonucleotide, underlined); these sites are needed for the cloning procedure.

The amplified DNA fragment was cloned into a pCRII vector and called pCRII-TIMP1. This vector was subsequently digested with the restriction enzymes NruI and BsteII and the TIMP1 containing DNA fragment was cloned into the pCRII-ATF plasmid (see figure 1).

In a next step the ATF-TIMP sequence was cloned into pMAD5. This was done by digestion of the pCRII-ATF-TIMP plasmid with the restriction enzymes EcoRV and SpeI, isolation of the ATF-TIMP encoding DNA fragment and cloning of this fragment into the SnabI and SpeI digested pMAD5 plasmid. The cloning was tested by restriction analysis and sequence analysis.

For construction of a recombinant adenovirus containing sequences encoding the ATF.TIMP1 hybrid protein, this sequence is cloned in the pMAD5 expression plasmid. This plasmid contains part of the wildtype adenovirus type 5 DNA sequences, a Major Late Promoter (MLP), and a poly-adenylation (polyA) signal and can be used as either an expression vector or a shuttle vector to construct a recombinant adenovirus. To clone the ATF.TIMP1 sequence into the pMAD5 plasmid between the MLP promoter and the polyA signal the following strategy has been followed.

Starting from a pCRII plasmid in which a 1373 base pair fragment of the uPA cDNA was cloned, a PCR reaction with the oligonucleotides 5'-cccgggctttttccatctgcgcagtc-3' and 5'-agggtcaccaaggaagagaatggc-3' was performed as described in example 1 to make a pCRII-ATF plasmid (see figure 1). Subsequently on this pCRII-ATF plasmid a PCR reaction was performed using the oligonucleotides 5'-aatattattgaacttcatcaagttcc-3' and 5'-gactctagagcaaaaatgacaaccag-3' and the resulting DNA fragment was cloned into the pCRII cloning vector. In this way the signal peptide of u-PA is removed and a SspI

restriction enzyme recognition site is introduced (underlined). The resulting plasmid DNA is designated pCRIIATF*.

In parallel a fragment of the cDNA of TIMP1 in pUC13-TIMP1 encoding amino acid residues -23 to 184 of the TIMP-1 protein, including the signal peptide but lacking the stop codon, was amplified using the oligonucleotides 5'-agagagacaccagagaacccaccat-3' and 5'-aatattggctatctgggaccgcagg-3' containing a recognition site for the restriction enzyme Ssp1 (underlined) and cloned into a pCRII cloning vector. The resulting plasmid DNA is designated pCRII-TIMP1*.

This vector was subsequently digested with the restriction enzymes SspI and EcoRV and the TIMP1 containing DNA fragment was cloned into a EcoRV-SspI digested pCRII-ATF* plasmid. The resulting plasmid containing the TIMP-ATF DNA fragment was called pCRII-TIMP-ATF. In a next step, the TIMP-ATF sequence was cloned into pMAD5. This was done by digestion of the pCRII-TIMP-ATF plasmid with the restriction enzymes EcoRV and SpeI, isolation of the TIMP-ATF encoding DNA fragment and cloning of this fragment into the SnabI and SpeI digested pMAD5 plasmid. The cloning was tested by restriction analysis and sequence analysis.

EXAMPLE 7

Vectors encoding hybrid proteins containing multiple copies of the BPTI unit coupled to the ATF domain have been constructed. To construct these multiple BPTI vectors, the following strategy is followed.

The pMAD5-ATF-BPTI described in EXAMPLE 4 is digested with the restriction enzymes SspI and BstEII. In this way the vector is opened exactly in the open reading frame at the end of the BPTI sequence. The pCRII-BPTI plasmid described in EXAMPLE 2 is digested with NruI and BstEII resulting in a BPTI encoding DNA fragment with one blunt end (NruI). The fragment was then monodirectionally cloned into the SspI BstEII pMAD5-ATF-BPTI vector. The thus constructed

plasmid named pMAD5-ATF-BPTI-BPTI was used as a shuttle vector for the construction of recombinant adenoviruses.

This approach can be repeated multiple times to construct vectors containing multiple BPTI-domains.

EXAMPLE 8

A vector encoding a hybrid protein containing both a BPTI unit and a TIMP1 unit coupled to the ATF domain has been constructed. To construct this BPTI-TIMP vector, the following strategy is followed.

The pMAD5-ATF-BPTI described in EXAMPLE 4 is digested with the restriction enzymes SspI and BstEII. In this way the vector is opened right behind the BPTI sequence. The pCRII-TIMP plasmid described in EXAMPLE 6 is digested with NruI and BstEII resulting in a TIMP1 encoding DNA fragment with one blunt end. The fragment was then cloned into the SspI BstEII pMAD5-ATF-BPTI vector. The thus constructed plasmid named pMAD5-ATF-BPTI-TIMP was used as a shuttle vector for the construction of recombinant adenoviruses.

EXAMPLE 9

To monitor the production of a functional ATF-BPTI hybrid protein after transfection of cells with pMAD5 or transduction with a recombinant replication-deficient ATF-BPTI encoding adenovirus, the following tests have been performed.

- The production of the hybrid ATF-BPTI protein by CHO cells transfected with the pMAD5-ATF-BPTI was analyzed using a uPA ELISA that recognizes the ATF, the aminoterminal fragment of u-PA. Production of ATF-BPTI was clearly detectable both after transient transfection of CHO cells with the pMAD5-ATF-BPTI plasmid (50-100 ng/ml/24hrs) and after transduction with an ATF-BPTI encoding adenoviral vector (up to 1.5 µg/ml/24hrs).
- The cell culture media of CHO cells transduced with the ATF-BPTI adenovirus were analyzed using western blotting

techniques. After electrophoresis and blotting, parallel filters were analyzed with polyclonal antibodies against either u-PA or BPTI (raised against Trasylol®). In both filters a signal was detected at the same expected position at approximately 20kDa. This indicates that the protein produced indeed contains fragments of u-PA and BPTI, thus that the hybrid protein is produced.

- The function as an inhibitor of plasmin activity of the ATF-BPTI protein was first analyzed in solution using dilutions of the culture medium of ATF-BPTI virus infected CHO cells (approximately 1.8 µg/ml). They were incubated with plasmin (1 nM) and the activity of plasmin was measured using a chromogenic substrate. Trasylol® dilutions were used as control references. Plasmin inhibition by ATF-BPTI medium was very effective, diluting the medium 1000x (i.e. 100 nM ATF-BPTI) resulted in a 50% inhibition of the activity of 1 nM plasmin, a similar inhibition as was observed with 100 nM Trasylol®. Thus the activity of ATF-BPTI is comparable to that of commercially available Trasylol® (Bayer, Germany).

- The function of ATF-BPTI as an inhibitor for plasmin bound to the cell surface via the interaction of the ATF domain with the u-PA receptor (uPAR) was tested using mouse cell lines that are either or not transfected with the human uPA receptor gene. These cells were incubated for 6 hrs with diluted medium of the ATF-BPTI virus-infected CHO cells. Cell extracts were made of the uPAR-transfected cells and the parental mouse cells lacking the human uPAR. Parallel cultures underwent a short acid treatment (pH 3, 3 min) before the cell extracts were made. This treatment will remove any u-PA or ATF bound to the u-PA receptor. The cell extracts were incubated with 1nM plasmin and the plasmin activity was determined. Plasmin activity could only be inhibited by the cell extract of the u-PAR containing cell line. No inhibition of plasmin activity was observed in the cell extracts of parental cell line, lacking the u-PA receptor, and in the acid-treated u-PAR containing cell line.

This clearly indicates that ATF-BPTI can function as a u-PAR bound plasmin inhibitor.

TABLE 1

	% INHIBITION OF PLASMIN ACTIVITY			
cell line	uPAR transfected cell line		parental cell line	
acid treatment	-	+	-	+
% inhibition	93%	0%	0%	0%

EXAMPLE 10

Cell-specific expression of ATF-BPTI in endothelial cells e.g. to specifically inhibit the migration of endothelial cells during angiogenesis, is achieved by cloning sequences of the promoter of the human pre-pro-endothelin 1 gene (nucleotide 2180-3680 of HUMEDN1B (GENBANK)) in front of the ATF-BPTI encoding DNA in an adenoviral vector. In this way, highly endothelial cell-specific expression of the ATF-BPTI hybrid protein can be obtained.

EXAMPLE 11

Proteolytic degradation of the extracellular matrix is a key event in many cell migration and tissue remodeling processes. This proteolytic matrix degradation is often found to be mediated by urokinase-type plasminogen activation. In order to test whether infection with an ATF-BPTI encoding adenovirus can inhibit plasmin mediated extracellular matrix degradation, an experiment was performed using human synoviocytes. These cells were infected with an ATF-BPTI adenovirus while they were seeded on an ^{3}H-labeled extracellular matrix existing of bovine cartilage material. Profound inhibition of matrix degradation could be observed in the virus treated cells (figure 4) indicating that matrix

degradation can be inhibited by infecting cells with the ATF-BPTI encoding virus.

Figure 4 shows the degradation of cartilage matrix by human synoviocytes in the presence of plasminogen. Matrix is incubated with control medium (lane 1), synoviocytes (lane 2), synoviocytes infected with ATF-BPTI adenovirus (lane 3), and synoviocytes incubated with Trasylol®(100KIU/ml)(lane 4).

EXAMPLE 12

In the process of restenosis smooth muscle cell migration and vessel wall remodeling are key events in which plasmin mediated proteolysis of extracellular matrix components is involved. In vivo application of general plasmin inhibitors to interfere in this process may have systemic side effects. Application of a plasmin inhibitor to the surface of the migrating cells might prevent these side effects. Infection of the blood vessel wall with an ATF-BPTI adenovirus at a site where neointima formation can be expected, e.g. in a transplanted "coronary by-pass" graft, might be a ideal way to produce the ATF-BPTI locally, and thus inhibit plasmin activity in the direct surroundings of the migrating (smooth muscle) cells, resulting in a reduced neointima formation.

This principle was tested using human saphenous vein organ cultures, a model system in which neointima formation can be mimicked very realistically. In parallel cultures, either or not infected with an ATF-BPTI adenovirus, the neointima formation was analyzed after three and four weeks. In the untreated tissues a clear neointima formation could be observed. Profound inhibition of the neointima formation could be observed in the tissues treated with 10^{10} pfu/ml ATF-BPTI adenovirus.

Appendix

Description and Nucleotide sequence of the pMAD5-ATF-BPTI plasmid.

From	To	Description
1	184	adenovirus sequence 5'
184	447	adenovirus Major Late Promoter (MLP)
447	644	tripartite leader sequence (TPL)
685	1167	urokinase ATF sequence
1168	1353	bovine prancreas trypsin inhibitor sequence
1360	1443	urokinase 3' sequence (including stop codon)
1514	1615	sequence derived form pSP65 and LacZ
1616	1751	SV40 poly A signal
1752	7334	adenovirus sequence 3'
9831	8971	β-lactamase

Nucleotide sequence:

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1     CATTTTCGCG GGAAAACTGA ATAAGAGGAA GTGAAATCTG AATAATTTTG TGTTACTCAT
61    AGCGCGTAAT ATTTGTCTAG GGCCGCGGGG ACTTTGACCG TTTACGTGGA GACTCGCCCA
121   GGTGTTTTTC TCAGGTGTTT TCCGCGTTCC GGGTCAAAGT TGGCGTTTTA TTATTATAGT
181   CAGCTGATCG AGCGGTGTTC CGCGGTCCTC CTCGTATAGA AACTCGGACC ACTCTGAGAC
241   GAAGGCTCGC GTCCAGGCCA GCACGAAGGA GGCTAAGTGG GAGGGGTAGC GGTCGTTGTC
301   CACTAGGGGG TCCACTCGCT CCAGGGTGTG AAGACACATG TCGCCCTCTT CGGCATCAAG
361   GAAGGTGATT GGTTTATAGG TGTAGGCCAC GTGACCGGGT GTTCCTGAAG GGGGGCTATA
421   AAAGGGGGTG GGGGCGCGTT CGTCCTCACT CTCTTCCGCA TCGCTGTCTG CGAGGGCCAG
481   CTGTTGGGGC TCGCGGTTGA GGACAAACTC TTCGCGGTCT TTCCAGTACT CTTGGATCGG
541   AAACCCGTCG GCCTCCGAAC GGTACTCCGC CACCGAGGGA CCTGAGCGAG TCCGCATCGA
601   CCGGATCGGA AAACCTCTCG AGAAAGGCGT CTAACCAGTC GCTGATCGAT AAGCTAGCTT
661   ACGCGTACAT CTGCAGAATT CGGCTTAACT CTAGACCATG AGAGCCCTGC TGGCGCGCCT
721   GCTTCTCTGC GTCCTGGTCG TGAGCGACTC CAAAGGCAGC AATGAACTTC ATCAAGTTCC
781   ATCGAACTGT GACTGTCTAA ATGGAGGAAC ATGTGTGTCC AACAAGTACT TCTCCAACAT
841   TCACTGGTGC AACTGCCCAA AGAAATTCGG AGGGCAGCAC TGTGAAATAG ATAAGTCAAA
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901  AACCTGCTAT GAGGGGAATG GTCACTTTTA CCGAGGAAAG GCCAGCACTG ACACCATGGG
961  CCGGCCCTGC CTGCCCTGGA ACTCTGCCAC TGTCCTTCAG CAAACGTACC ATGCCCACAG
1021 ATCTGATGCT CTTCAGCTGG GCCTGGGGAA ACATAATTAC TGCAGGAACC CAGACAACCG
1081 GAGGCGACCC TGGTGCTATG TGCAGGTGGG CCTAAAGCCG CTTGTCCAAG AGTGCATGGT
1141 GCATGACTGC GCAGATGGAA AAAAGCCCCG ACCTGACTTC TGCCTAGAGC CTCCATATAC
1201 GGGTCCCTGC AAGGCCAGAA TTATCAGATA CTTCTACAAC GCCAAGGCTG GGCTCTGCCA
1261 GACCTTTGTA TATGGCGGCT GCAGAGCTAA AAGAAACAAT TTCAAGAGCG CAGAGGACTG
1321 CATGAGGACC TGTGGTGGTA ATATTGGGCC CTGGGTCACC AAGGAAGAGA ATGGCCTGGC
1381 CCTCTGAGGG TCCCCAGGGA GGAAACGGGC ACCACCCGCT TTCTTGCTGG TTGTCATTTT
1441 TGCTCTAGAG TCAAGCCGAA TTCTGCAGAT ATCGTCCATT CCGACAGCAT CGCCAGTCAC
1501 TATGGCGTGC TGCTAGAGGA TCCCCGGGCG AGCTCGAATT CCAGCTGAGC GCCGGTCGCT
1561 ACCATTACCA GTTGGTCTGG TGTCAAAAAT AATAATAACC GGGCAGGGGG GATTCTGAAC
1621 TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA GCATCACAAA TTTCACAAAT
1681 AAAGCATTTT TTTCACTGCA TTCTAGTTGT GGTTTGTCCA AACTCATCAA TGTATCTTAT
1741 CATGTCTGGA TCTGGAAGGT GCTGAGGTAC GATGAGACCC GCACCAGGTG CAGACCCTGC
1801 GAGTGTGGCG GTAAACATAT TAGGAACCAG CCTGTGATGC TGGATGTGAC CGAGGAGCTG
1861 AGGCCCGATC ACTTGGTGCT GGCCTGCACC CGCGCTGAGT TTGGCTCTAG CGATGAAGAT
1921 ACAGATTGAG GTACTGAAAT GTGTGGGCGT GGCTTAAGGG TGGGAAAGAA TATATAAGGT
1981 GGGGGTCTTA TGTAGTTTTG TATCTGTTTT GCAGCAGCCG CCGCCGCCAT GAGCACCAAC
2041 TCGTTTGATG GAAGCATTGT GAGCTCATAT TTGACAACGC GCATGCCCCC ATGGGCCGGG
2101 GTGCGTCAGA ATGTGATGGG CTCCAGCATT GATGGTCGCC CCGTCCTGCC CGCAAACTCT
2161 ACTACCTTGA CCTACGAGAC CGTGTCTGGA ACGCCGTTGG AGACTGCAGC CTCCGCCGCC
2221 GCTTCAGCCG CTGCAGCCAC CGCCCGCGGG ATTGTGACTG ACTTTGCTTT CCTGAGCCCG
2281 CTTGCAAGCA GTGCAGCTTC CCGTTCATCC GCCCGCGATG ACAAGTTGAC GGCTCTTTTG
2341 GCACAATTGG ATTCTTTGAC CCGGGAACTT AATGTCGTTT CTCAGCAGCT GTTGGATCTG
2401 CGCCAGCAGG TTTCTGCCCT GAAGGCTTCC TCCCCTCCCA ATGCGGTTTA AAACATAAAT
2461 AAAAAACCAG ACTCTGTTTG GATTTGGATC AAGCAAGTGT CTTGCTGTCT TTATTTAGGG
2521 GTTTTGCGCG CGCGGTAGGC CCGGGACCAG CGGTCTCGGT CGTTGAGGGT CCTGTGTATT
2581 TTTTCCAGGA CGTGGTAAAG GTGACTCTGG ATGTTCAGAT ACATGGGCAT AAGCCCGTCT
2641 CTGGGGTGGA GGTAGCACCA CTGCAGAGCT TCATGCTGCG GGGTGGTGTT GTAGATGATC
2701 CAGTCGTAGC AGGAGCGCTG GGCGTGGTGC CTAAAAATGT CTTTCAGTAG CAAGCTGATT
2761 GCCAGGGGCA GGCCCTTGGT GTAAGTGTTT ACAAAGCGGT TAAGCTGGGA TGGGTGCATA
2821 CGTGGGGATA TGAGATGCAT CTTGGACTGT ATTTTTAGGT TGGCTATGTT CCCAGCCATA
2881 TCCCTCCGGG GATTCATGTT GTGCAGAACC ACCAGCACAG TGTATCCGGT GCACTTGGGA
2941 AATTTGTCAT GTAGCTTAGA AGGAAATGCG TGGAAGAACT TGGAGACGCC CTTGTGACCT
3001 CCAAGATTTT CCATGCATTC GTCCATAATG ATGGCAATGG GCCCACGGGC GGCGGCCTGG
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3061  GCGAAGATAT TTCTGGGATC ACTAACGTCA TAGTTGTGTT CCAGGATGAG ATCGTCATAG
3121  GCCATTTTTA CAAAGCGCGG GCGGAGGGTG CCAGACTGCG GTATAATGGT TCCATCCGGC
3181  CCAGGGGCGT AGTTACCCTC ACAGATTTGC ATTTCCCACG CTTTGAGTTC AGATGGGGGG
3241  ATCATGTCTA CCTGCGGGGC GATGAAGAAA ACGGTTTCCG GGGTAGGGGA GATCAGCTGG
3301  GAAGAAAGCA GGTTCCTGAG CAGCTGCGAC TTACCGCAGC CGGTGGGCCC GTAAATCACA
3361  CCTATTACCG GGTGCAACTG GTAGTTAAGA GAGCTGCAGC TGCCGTCATC CCTGAGCAGG
3421  GGGGCCACTT CGTTAAGCAT GTCCCTGACT CGCATGTTTT CCCTGACCAA ATCCGCCAGA
3481  AGGCGCTCGC CGCCCAGCGA TAGCAGTTCT TGCAAGGAAG CAAAGTTTTT CAACGGTTTG
3541  AGACCGTCCG CCGTAGGCAT GCTTTTGAGC GTTTGACCAA GCAGTTCCAG GCGGTCCCAC
3601  AGCTCGGTCA CCTGCTCTAC GGCATCTCGA TCCAGCATAT CTCCTCGTTT CGCGGGTTGG
3661  GGCGGCTTTC GCTGTACGGC AGTAGTCGGT GCTCGTCCAG ACGGGCCAGG GTCATGTCTT
3721  TCCACGGGCG CAGGGTCCTC GTCAGCGTAG TCTGGGTCAC GGTGAAGGGG TGCGCTCCGG
3781  GCTGCGCGCT GGCCAGGGTG CGCTTGAGGC TGGTCCTGCT GGTGCTGAAG CGCTGCCGGT
3841  CTTCGCCCTG CGCGTCGGCC AGGTAGCATT TGACCATGGT GTCATAGTCC AGCCCCTCCG
3901  CGGCGTGGCC CTTGGCGCGC AGCTTGCCCT TGGAGGAGGC GCCGCACGAG GGGCAGTGCA
3961  GACTTTTGAG GGCGTAGAGC TTGGGCGCGA GAAATACCGA TTCCGGGGAG TAGGCATCCG
4021  CGCCGCAGGC CCCGCAGACG GTCTCGCATT CCACGAGCCA GGTGAGCTCT GGCCGTTCGG
4081  GGTCAAAAAC CAGGTTTCCC CCATGCTTTT TGATGCGTTT CTTACCTCTG GTTTCCATGA
4141  GCCGGTGTCC ACGCTCGGTG ACGAAAAGGC TGTCCGTGTC CCCGTATACA GACTTGAGAG
4201  GCCTGTCCTC GAGCGGTGTT CCGCGGTCCT CCTCGTATAG AAACTCGGAC CACTCTGAGA
4261  CAAAGGCTCG CGTCCAGGCC AGCACGAAGG AGGCTAAGTG GGAGGGGTAG CGGTCGTTGT
4321  CCACTAGGGG GTCCACTCGC TCCAGGGTGT GAAGACACAT GTCGCCCTCT TCGGCATCAA
4381  GGAAGGTGAT TGGTTTGTAG GTGTAGGCCA CGTGACCGGG TGTTCCTGAA GGGGGGCTAT
4441  AAAAGGGGGT GGGGGCGCGT TCGTCCTCAC TCTCTTCCGC ATCGCTGTCT GCGAGGGCCA
4501  GCTGTTGGGG TGAGTACTCC CTCTGAAAAG CGGGCATGAC TTCTGCGCTA AGATTGTCAG
4561  TTTCCAAAAA CGAGGAGGAT TTGATATTCA CCTGGCCCGC GGTGATGCCT TTGAGGGTGG
4621  CCGCATCCAT CTGGTCAGAA AAGACAATCT TTTTGTTGTC AAGCTTGGTG GCAAACGACC
4681  CGTAGAGGGC GTTGGACAGC AACTTGGCGA TGGAGCGCAG GGTTTGGTTT TTGTCGCGAT
4741  CGGCGCGCTC CTTGGCCGCG ATGTTTAGCT GCACGTATTC GCGCGCAACG CACCGCCATT
4801  CGGGAAAGAC GGTGGTGCGC TCGTCGGGCA CCAGGTGCAC GCGCCAACCG CGGTTGTGCA
4861  GGGTGACAAG GTCAACGCTG GTGGCTACCT CTCCGCGTAG GCGCTCGTTG GTCCAGCAGA
4921  GGCGGCCGCC CTTGCGCGAG CAGAATGGCG GTAGGGGGTC TAGCTGCGTC TCGTCCGGGG
4981  GGTCTGCGTC CACGGTAAAG ACCCCGGGCA GCAGGCGCGC GTCGAAGTAG TCTATCTTGC
5041  ATCCTTGCAA GTCTAGCGCC TGCTGCCATG CGCGGGCGGC AAGCGCGCGC TCGTATGGGT
5101  TGAGTGGGGG ACCCCATGGC ATGGGGTGGG TGAGCGCGGA GGCGTACATG CCGCAAATGT
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5761  TGTCGTCGCA TCCGCCCTGC TCCCAGAGCA AAAAGTCCGT GCGCTTTTTG GAACGCGGAT
5821  TTGGCAGGGC GAAGGTGACA TCGTTGAAGA GTATCTTTCC CGCGCGAGGC ATAAAGTTGC
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5941  CGATCTCGTC AAAGCCGTTG ATGTTGTGGC CCACAATGTA AAGTTCCAAG AAGCGCGGGA
6001  TGCCCTTGAT GGAAGGCAAT TTTTTAAGTT CCTCGTAGGT GAGCTCTTCA GGGGAGCTGA
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6781  TGTCCATGGT CTGGAGCTCC CGCGGCGTCA GGTCAGGCGG GAGCTCCTGC AGGTTTACCT
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7201  GAGCCTGAAA GAGAGTTCGA CAGAATCAAT TTCGGTGTCG TTGACGGCGG CCTGGCGCAA
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7321  CTCTTCCTCC TGGAGATCAA TTGAAGCTAG CTTTAATGCG GTAGTTTATC ACAGTTAAAT
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7501  CGGGATATCG TCCATTCCGA CAGCATCGCC AGTCACTATG GCGTGCTGCT AGCGCTATAT
7561  GCGTTGATGC AATTTCTATG CGCACCCGTT CTCGGAGCAC TGTCCGACCG CTTTGGCCGC
7621  CGCCCAGTCC TGCTCGCTTC GCTACTTGGA GCCACTATCG ACTACGCGAT CATGGCGACC
7681  ACACCCGTCC TGTGGATCTC GACCGATGCC CTTGAGAGCC TTCAACCCAG TCAGCTCCTT
7741  CCGGTGGGCG CGGGGCATGA CTATCGTCGC CGCACTTATG ACTGTCTTCT TTATCATGCA
7801  ACTCGTAGGA CAGGTGCCGG CAGCGCTCTG GGTCATTTTC GGCGAGGACC GCTTTCGCTG
7861  GAGCGCGACG ATGATCGGCC TGTCGCTTGC GGTATTCGGA ATCTTGCACG CCCTCGCTCA
7921  AGCCTTCGTC ACTGGTCCCG CCACCAAACG TTTCGGCGAG AAGCAGGCCA TTATCGCCGG
7981  CATGGCGGCC GACGCGCTGG GCTACGTCTT GCTGGCGTTC GCGACGCGAG GCTGGATGGC
8041  CTTCCCCATT ATGATTCTTC TCGCTTCCGG CGGCATCGGG ATGCCCGCGT TGCAGGCCAT
8101  GCTGTCCAGG CAGGTAGATG ACGACCATCA GGGACAGCTT CAAGGATCGC TCGCGGCTCT
8161  TACCAGCCCA GCAAAAGGCC AGGAACCGTA AAAAGGCCGC GTTGCTGGCG TTTTTCCATA
8221  GGCTCCGCCC CCCTGACGAG CATCACAAAA ATCGACGCTC AAGTCAGAGG TGGCGAAACC
8281  CGACAGGACT ATAAAGATAC CAGGCGTTTC CCCCTGGAAG CTCCCTCGTG CGCTCTCCTG
8341  TTCCGACCCT GCCGCTTACC GGATACCTGT CCGCCTTTCT CCCTTCGGGA AGCGTGGCGC
8401  TTTCTCATAG CTCACGCTGT AGGTATCTCA GTTCGGTGTA GGTCGTTCGC TCCAAGCTGG
8461  GCTGTGTGCA CGAACCCCCC GTTCAGCCCG ACCGCTGCGC CTTATCCGGT AACTATCGTC
8521  TTGAGTCCAA CCCGGTAAGA CACGACTTAT CGCCACTGGC AGCAGCCACT GGTAACAGGA
8581  TTAGCAGAGC GAGGTATGTA GGCGGTGCTA CAGAGTTCTT GAAGTGGTGG CCTAACTACG
8641  GCTACACTAG AAGGACAGTA TTTGGTATCT GCGCTCTGCT GAAGCCAGTT ACCTTCGGAA
8701  AAAGAGTTGG TAGCTCTTGA TCCGGCAAAC AAACCACCGC TGGTAGCGGT GGTTTTTTTG
8761  TTTGCAAGCA GCAGATTACG CGCAGAAAAA AAGGATCTCA AGAAGATCCT TTGATCTTTT
8821  CTACGGGGTC TGACGCTCAG TGGAACGAAA ACTCACGTTA AGGGATTTTG GTCATGAGAT
8881  TATCAAAAAG GATCTTCACC TAGATCCTTT TAAATTAAAA ATGAAGTTTT AAATCAATCT
8941  AAAGTATATA TGAGTAAACT TGGTCTGACA GTTACCAATG CTTAATCAGT GAGGCACCTA
9001  TCTCAGCGAT CTGTCTATTT CGTTCATCCA TAGTTGCCTG ACTCCCCGTC GTGTAGATAA
9061  CTACGATACG GGAGGGCTTA CCATCTGGCC CCAGTGCTGC AATGATACCG CGAGACCCAC
9121  GCTCACCGGC TCCAGATTTA TCAGCAATAA ACCAGCCAGC CGGAAGGGCC GAGCGCAGAA
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9301  TGTCACGCTC GTCGTTTGGT ATGGCTTCAT TCAGCTCCGG TTCCCAACGA TCAAGGCGAG
9361  TTACATGATC CCCCATGTTG TGCAAAAAAG CGGTTAGCTC CTTCGGTCCT CCGATCGTTG
9421  TCAGAAGTAA GTTGGCCGCA GTGTTATCAC TCATGGTTAT GGCAGCACTG CATAATTCTC
9481  TTACTGTCAT GCCATCCGTA AGATGCTTTT CTGTGACTGG TGAGTACTCA ACCAAGTCAT
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9601  CCGCGCCACA TAGCAGAACT TTAAAAGTGC TCATCATTGG AAAACGTTCT TCGGGGCGAA
9661  AACTCTCAAG GATCTTACCG CTGTTGAGAT CCAGTTCGAT GTAACCCACT CGTGCACCCA
9721  ACTGATCTTC AGCATCTTTT ACTTTCACCA GCGTTTCTGG GTGAGCAAAA ACAGGAAGGC
9781  AAAATGCCGC AAAAAAGGGA ATAAGGGCGA CACGGAAATG TTGAATACTC ATACTCTTCC
9841  TTTTTCAATA TTATTGAAGC ATTTATCAGG GTTATTGTCT CATGAGCGGA TACATATTTG
9901  AATGTATTTA GAAAAATAAA CAAATAGGGG TTCCGCGCAC ATTTCCCCGA AAAGTGCCAC
9961  CTGACGTCTA AGAAACCATT ATTATCATGA CATTAACCTA TAAAAATAGG CGTATCACGA
10021 GGCCCTTTCG TCTTCAAGAA TTCTCATGTT TGACAGCTTA TCATCATCAA TAATATACCT
10081 TATTTTGGAT TGAAGCCAAT ATGATAATGA GGGGGTGGAG TTTGTGACGT GGCGCGGGGC
10141 GTGGGAACGG GGCGGGTGAC GTAGTAGTGT GGCGGAAGTG TGATGTTGCA AGTGTGGCGG
10201 AACACATGTA AGCGACGGAT GTGGCAAAAG TGACGTTTTT GGTGTGCGCC GGTGTACACA
10261 GGAAGTGACA ATTTTCGCGC GGTTTTAGGC GGATGTTGTA GTAAATTTGG GCGTAACCGA
10321 GTAAGATTTG GC
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Claims

1. A recombinant nucleic acid molecule comprising a vector useful for transfection or transduction of mammalian, e.g. human, cells, wherein said vector contains a nucleic acid insertion encoding an expressible hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function.

2. A recombinant nucleic acid molecule according to Claim 1, wherein said domain with a binding function comprises a receptor binding domain.

3. A recombinant nucleic acid molecule according to Claim 2, wherein said receptor binding domain is selected from the group consisting of urokinase receptor binding domain of urokinase, receptor binding domain of epidermal growth factor, receptor associated protein that binds to LDL Receptor related protein (α_2-macroglobulin receptor) and VLDL Receptor.

4. A recombinant nucleic acid molecule according to Claim 2, wherein said receptor binding domain comprises the aminoterminal part of urokinase which is capable of binding to the urokinase receptor.

5. A recombinant nucleic acid molecule according to Claim 2, wherein said receptor binding domain comprises amino acid residues 1 through 135 of urokinase.

6. A recombinant nucleic acid molecule according to Claim 1, wherein said domain with an effector function is an enzymatically active domain.

7. A recombinant nucleic acid molecule according to Claim 1, wherein said domain with an effector function has protease inhibitor activity.

8. A recombinant nucleic acid molecule according to Claim 7, wherein said domain having protease inhibitor activity comprises a protease inhibitor or active part

thereof, said protease inhibitor being selected from the group consisting of (bovine) pancreatic trypsin inhibitor, (bovine) splenic trypsin inhibitor, urinary trypsin inhibitor, tissue inhibitor of matrix metalloproteinase 1, tissue inhibitor of matrix metalloproteinase 2, tissue inhibitor of matrix metalloproteinase 3, and elastase inhibitor.

9. A recombinant nucleic acid molecule according to Claim 7, wherein said domain having protease inhibitor activity comprises (amino acid residues 53 through 94 of) mature bovine pancreatic trypsin inhibitor.

10. A recombinant nucleic acid molecule according to Claim 7, wherein said domain having protease inhibitor activity comprises bovine splenic trypsin inhibitor.

11. A recombinant nucleic acid molecule according to Claim 7, wherein said domain having protease inhibitor activity comprises a tissue inhibitor of matrix metallo-proteinases.

12. A recombinant nucleic acid molecule according to Claim 1, wherein said domain with an effector function comprises (an active part of) two or more different protease inhibitors, or two or more copies of (an active part of) a protease inhibitor, or both.

13. A recombinant nucleic acid molecule according to Claim 1, wherein said vector is selected from the group consisting of viral and non-viral vectors useful for transfection or transduction of mammalian cells.

14. A recombinant nucleic acid molecule according to Claim 1, wherein said vector is an adenovirus vector or a retrovirus vector useful for transfection or transduction of human cells.

15. A recombinant nucleic acid molecule according to Claim 1, wherein said vector is an adenovirus vector based on shuttle vector pMAD5.

16. A recombinant nucleic acid molecule according to Claim 1, wherein said nucleic acid insertion encoding an

expressible hybrid polypeptide or protein is under the control of a cell- or tissue-specific promoter.

17. A recombinant nucleic acid molecule according to Claim 1, wherein said nucleic acid insertion encoding an expressible hybrid polypeptide or protein is under the control of an endothelial cell-specific promoter, or a vascular smooth muscle cell-specific promoter, or a liver-specific promoter.

18. A process for preventing local proteolytic activity, extracellular matrix degradation, cell migration, cell invasion, or tissue remodeling, comprising transfecting or transducing the cells involved or cells in their environment with a recombinant nucleic acid molecule as claimed in any one of the preceding Claims to obtain local expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule.

19. A process for producing a hybrid polypeptide or protein which comprises a domain with a binding function and a domain with an effector function, comprising transfecting or transducing mammalian cells with a recombinant nucleic acid molecule as claimed in any one of Claims 1 to 17 to obtain expression of the hybrid polypeptide or protein encoded by said nucleic acid molecule, and optionally recovering the hybrid polypeptide or protein produced.

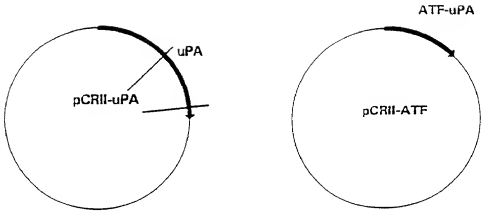
uPA
pCRII-uPA
ATF-uPA
pCRII-ATF

Fig. 1

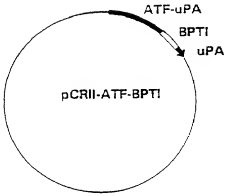
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Fig. 2

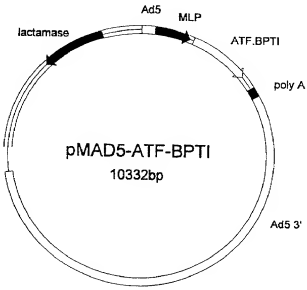

Fig. 3

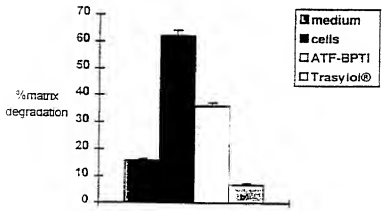

Fig. 4

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FULL NAME OF 6TH INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME
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I further declare that all statements made herein of my own knowledge are true, and that all statements on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

1st Inventor _____ Date 15-11-99

2nd Inventor _____ Date 15-11-99

3rd Inventor _____ Date _____

4th Inventor _____ Date _____

5th Inventor _____ Date _____

6th Inventor _____ Date _____

Rev. 1-3/11/83

Effective February 27, 1983

DECLARATION AND POWER OF ATTORNEY FOR U.S. PATENT APPLICATIONS

( ) Original ( ) Supplemental ( ) Substitute (X) PCT

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated below next to my name; that I verily believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Title: Method and construct for inhibition of cell migration

which is described and claimed in:

( ) the attached specification, or

(X) the specification in the application Serial No. _____ filed 12 November 1999 ; and with amendments through _____ (if applicable),

(X) the specification in International Application No. PCT/NL98/00259 _____, filed 11 May 1998 , and as amended on _____ (if applicable).

I hereby state that I have reviewed and understand the content of the above-identified specification, including the claims, as amended by any amendment(s) referred to above.

I acknowledge my duty to disclose information of which I am aware which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, §1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

COUNTRY	APPLICATION NO.	DATE OF FILING	PRIORITY CLAIMED
EP	97201423.7	12 May 1997	(X) YES ( ) NO
			( ) YES ( ) NO
			( ) YES ( ) NO
			( ) YES ( ) NO
			( ) YES ( ) NO
			( ) YES ( ) NO

I hereby claim the benefit under Title 35, United States Code, §120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

SERIAL NO.	U.S. FILING DATE	STATUS
		( ) Patented ( ) Pending ( ) Abandoned
		( ) Patented ( ) Pending ( ) Abandoned
		( ) Patented ( ) Pending ( ) Abandoned

As a named inventor I hereby appoint the following attorney(s) and/or patent agent(s) to prosecute this application and to transact all business in the United States Patent and Trademark Office connected therewith: Bruce S. Londa, Reg. No. 33,531, Brian. L. Wamsley, Reg. No. 33, 045 and Alex L. Yip, Reg. No. 34,759.

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